OFFICIGENIN, A NEW SAPOGENIN OF GUAIACUM OFFICINALE

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ABSTRACT.—A new sapogenin, named officigenin (1), has been isolated from the acidic hydrolysate of the *Guaiacum officinale* saponins. The alkaline hydrolysis of 1 furnished two triterpenoids, namely 3β ,29-dihydroxyolean-12-en-28-oic acid (mesembryanthemoidigenic acid) (2) and 3β ,24-dihydroxyolean-12-en-28,29-dioic acid (3). Compounds 1 and 3 are new.

Recently, we reported (1) the isolation and characterization of a new sapogenin, namely 3β , 20ξ -dihydroxy-30-norolean-12-en-28-oic acid, and the artifacts 3β -hydroxy-30-norolean-12, 19-dien-28-oic acid and its methyl ester together with larreagenin A, β -sitosterol, and oleanolic acid, from the acid hydrolysate of the saponins obtained from the stem bark of *Guaiacum officinale* L. (Zygophyllaceae). We have now succeeded in isolating another sapogenin named officigenin from the same source.

Officigenin (1) in the acid hydrolysate was eluted with CHCl₃-MeOH (90:10) from the silica gel column (1) and purified through recrystallization from Me_2CO . It analyzed for C60H92O9 H2O, and its uv spectrum in MeOH showed only end absorption at 212 nm. The ir spectrum revealed the presence of hydroxyl (3440 cm^{-1}) , ester (1728 cm^{-1}) , carboxyl (1705 cm^{-1}) , and trisubstituted double bond(s) (825 cm^{-1}) . On treatment with CH_2N_2 , it furnished a dimethyl ester (1a), which was acetylated with Ac₂O and pyridine to give a dimethyl ester triacetate (**1b**). The ¹H-nmr spectrum of **1a** exhibited singlets due to 11 tertiary methyl groups at δ 0.66, 0.69, 0.75, 0.83, 0.88 (each 3H, s, $5 \times CH_3$), δ 0.96, 1.09, 1.22 (each 6H, s, $6 \times CH_3$); other ¹H-nmr signals were an unresolved double doublet centered at δ 2.88 (2H), a multiplet centered at δ 3.25, a singlet at δ 3.61 (6H) due to two COOMe, a singlet at δ 3.69 (2H), and a triplet at δ 5.28 (2H). From the ir and ¹H-nmr spectra, **1** appeared to be an ester of two triterpenoids, a carboxylic group of one esterified with an alcohol group of the other. This was also confirmed by alkaline hydrolysis of $\mathbf{1}$, which, indeed, afforded a mixture of two triterpenoids. These were separated through silica gel column chromatography yielding 3β , 29-dihydroxyolean-12-en-28-oic acid (mesembryanthemoidigenic acid) (2) and 3β , 24-dihydroxyolean-12-en-28, 29-dioic acid (3).

Compound 2 was eluted with C_6H_6 -EtOAc (1:1) and crystallized from MeOH as colorless needles. Its mass spectrum exhibited a molecular ion peak at m/z 472 corresponding to the formula $C_{30}H_{48}O_4$. On treatment with CH_2N_2 , it furnished a monoester (2a) which, on acetylation, gave a monoester diacetate (2b).





From ¹H-nmr and ms data the structure of **2** was deduced to be 3β , 29-dihydroxyolean-12-en-28-oic acid, which has previously been isolated from *Rhipsalis mesembryanthemoides* and named mesembryanthemoidigenic acid (2). This structure was also supported from the ¹³C-nmr spectrum of **2a** in CDCl₃ (Table 1). Peaks at 78.92, 122.69, and 143.44 ppm are due to C-3, C-12, and C-13, respectively. A peak at 74.33 ppm is ascribed to C-29. This peak is inverted in the gated spin echo (GASPE) spectrum, indicating that this peak is due to a methylene carbon bearing a primary OH group.



7 m/z=233 (37.07%)

Compound **3** was isolated from the fractions eluted with C_6H_6 -EtOAc (1:4), which crystallized from MeOH as small colorless crystals. Its mass spectrum showed a molecular ion peak at m/z 502, corresponding to the formula $C_{30}H_{46}O_6$. Its ir spectrum in KBr revealed the presence of hydroxyl (3425 cm⁻¹), carboxyl (1692 cm⁻¹), and trisubstitute double bond(s) (825 cm⁻¹). Its uv spectrum had a maximum at 210 nm (end absorption). Methylation with CH_2N_2 afforded a dimethyl ester (**3a**). The ¹H-nmr spectrum of **3a** showed five tertiary methyl signals at δ 0.66, 0.83, 1.10 (each 3H, s, 26, 25 and 27-Me groups, respectively), 1.22 (6H, s, 23 and 30-Me), a single proton double doublet centered at 2.88 (J=5, 13.75 Hz) characteristic of the H-18 proton of an olean-12-en-system (3), a multiplet centered at δ 3.35 attributed to H-3 and H-24, two carbomethoxy signals at δ 3.60 and 3.65, and a triplet at δ 5.29 (1H, J=3.75 Hz) ascribable to the proton at H-12, which is commonly encountered in the triterpenes of the oleanene and ursene series. Acetylation of **3a** with Ac₂O and pyridine furnished a

dimethyl ester diacetate (**3b**). Its ir spectrum showed no OH absorption. The mass spectrum of **3** revealed the characteristic fragmentation of the ring C of Δ^{12} amyrin derivatives with a base peak at m/z 278 and a peak at m/z 223 corresponding to ions 10 and 11 due to RDA fragmentation (4). The former peak is shifted at 306 in **3a** and remains unchanged in **3b**, indicating the presence of two carboxyl groups in ring D/E and two hydroxyl groups in ring A/B.

The ¹H-nmr spectrum of **3b** showed two acetoxy methyl signals at δ 2.02 and 2.04; a triplet centered at δ 4.57 (1H, J=8 Hz) is due to the H-3 α . The chemical shift and coupling pattern of this proton suggested that one OH group is located at C-3 and oriented in the β configuration. The ¹H-nmr spectrum of **3b** also showed an AB quartet centered at δ 4.25 (J=12.5 Hz) due to two methylene protons on a carbon atom bearing a primary acetoxyl group attached to an asymmetric center C-4. The chemical shift attributed to the methylene proton is within the range for an axial CH₂OAc (5). These data establish the orientation of the two hydroxyl groups in compound **3** as 3 β and 24. In the ¹H-nmr spectrum of **3b**, the highest C-methyl signal (δ 0.71) appeared upfield from δ 0.775 and one of the two methyl ester signals is at δ 3.60, suggesting the presence of a C-28 carboxyl group in the compound (6). The possible point of attachment of the other carboxyl group is at C-20. The influence of substitution on methyl frequencies in the ¹H-nmr spectrum of 12-oleanene derivatives was applied in order to

Carbon number	1aª	2a	1a	3a	Carbon number	1aª	2a	1a	3a
1	38.74	38.74	38.53	38.40	17	46.76	46.96	46.44	46.41
2	27.61	27.60	27.61	27.54	18	40.31	40.42	40.13	40.09
3	78.92	78.92	80.59	80.82	19	37.04	40.05	39.80	39.64
4	38.26	38.40	42.60	42.14	20	34.62	35.74	42.60	42.71
5	55.35	55.18	55.90	55.80	21	29.66	28.22	31.22	31.15
6	18.38	18.28	18.38	18.37	22	31.37	31.48	31.22	31.15
7	32.69	32.61	32.91	32.82	23	28.13	28.05	19.37	19.33
8	39.33	39.25	39.33	39.26	24	15.60	15.52	64.40	64.44
9	47.66	47.59	47.66	47.63	25	15.28	15.25	15.84	15.82
10	37.04	37.01	36.74	36.71	26	16.82	16.79	16.71	16.69
11	23.44	23.37	23.13	23.08	27	26.04	25.95	26.04	25.83
12	122.94	122.69	123.23	123.15	28	178.02	178.03	177.65	177.68
13	143.22	143.44	142.72	142.75	29	74.18	74.33	177.91	178.77
14	41.54	41.56	41.54	41.52	30	19.52	18.97	22.53	22.38
15	27.16	27.14	28.37	28.21	OMe	51.65	51.53	51.65	51.66
16	23.13	23.04	23.66	23.64	OMe			51.65	51.85

TABLE 1. ¹³C-nmr Chemical Shifts (in ppm) of Compounds 1a, 2a, and 3a in CDCl₃

^aThese values refer to the carbon numbers 1', 2' etc.

determine the orientation of the carboxyl group at C-20 (7) in **3**. The observed chemical shifts are in better agreement with those calculated for α carbomethoxy and β methyl groups attached to C-20 rather than β -carbomethoxy and α -methyl groups. Consequently, the structure and stereochemistry of **3** is established as 3β ,24-dihydroxy-olean-12-en-28,29-dioic acid. This proposed structure was also supported by the ¹³C-nmr spectrum of **3a** (Table 1), which showed signals for the double bond at 123.15 (C-12), 142.75 (C-13) ppm, close to the values in Δ^{12} oleanene series of triterpenoids. There were two carbon atoms bearing oxygen functions at 80.82 (C-3) and 64.44 ppm (C-24). The latter peak was inverted in the GASPE spectrum indicating that this carbon atom is a methylene, bearing a primary hydroxyl group.

The molecular formula of compound 1, $C_{60}H_{92}O_9$, suggested that this compound is an ester of compounds 2 and 3. In the ¹H-nmr spectrum of 1a the signal due to the 29-hydroxymethylene proton was shifted to relatively low field at δ 3.69 as compared to this signal in the ¹H-nmr of 2a at δ 3.25. This low field shift suggested that the 29hydroxymethylene proton of 2 is esterified with the carboxylic group of 3. On alkaline hydrolysis, 1 was easily and completely converted into 2 and 3. This ease of hydrolysis suggested that the 29-COOH (equatorial), rather than the more sterically hindered 28-COOH group, is involved in the ester formation. Thus, the ester linkage is between the 29-carboxylic group of 3 with the 29-hydroxyl group of 2. The ease of hydrolysis also confirmed the position of the carboxylic group in compound 3 at C-29, because a C-30 carboalkoxy group would not be easily hydrolyzed.

The low resolution mass spectrum of **1a** also supported the proposed structure. It showed a molecular ion peak at m/z 984.8 (calcd for $C_{62}H_{96}O_9$, 984.7). A peak of m/z 760.6 is due to structure **4** (calcd for $C_{48}H_{72}O_7$, 760.5). There is a peak at m/z 261 due to structure **6**. The base peak at m/z 201 arose due to the removal of HCOOCH₃ from **6**. The other RDA fragment, **5**, easily lost CO_2 , $(CO_2+COOCH_3)$, and $(CO_2+HCOOCH_3)$ giving rise to strong peaks at m/z 247, 188, and 187, respectively.

On the basis of the above spectra, structure **1** was suggested to be an ester of 3β , 24dihydroxyolean-12-en-28, 29-dioic acid (**3**) and 3β , 29-dihydroxyolean-12-en-28-oic



acid (2). This structure was also supported by a comparison of the 13 C-nmr spectral data of **1a** with those of **2a** and **3a** (Table 1).

In order to determine whether the ester of these two compounds (2 and 3) might have been formed during acidic hydrolysis of the saponins into sapogenins, a mixture of 2 and 3 was refluxed with methanolic HCl under the same conditions used for acidic hydrolysis of the crude saponins. It was found that mixtures of 2 and 3 do not yield the ester 1 under these conditions. This indicates that compound 1 is a genuine sapogenin and not an artifact formed during acidic hydrolysis of the saponins.

EXPERIMENTAL

GENERAL EXPERIMENTAL PROCEDURES.—The equipment used, the plant material, the isolation of saponins, acidic hydrolysis, and chromatography of the sapogenin mixture have been described in an earlier paper (1). The was carried out on silica gel plates using the following solvent system: (a) CHCl₃-MeOH (9:1), (b) C_6H_6 -EtOAc (9:1), and (c) C_6H_6 -EtOAc (7:3).

OFFICIGENIN (1).—Compound 1 was eluted from the silica gel column with CHCl₃-MeOH (9:1), crystallized from Me₂CO as colorless crystals (400 mg), mp, 264-265°, $[\alpha]D + 32.30$ (C=0.19, MeOH), uv λ max (MeOH) 212 nm (end absorption); ir ν max (KBr) 3440 (OH), 2940 (CH₂), 1728 (ester), 1705 (CO, acid), 1468, 1385, 1250, 1115, 1025, 995, 825 and 720 cm⁻¹. Anal. calcd for: C₆₀H₉₂O₉·H₂O; C, 73.88; H, 9.72. Found: C, 73.31; H, 9.54.

METHYLATION OF 1.—Compound 1 (100 mg) was dissolved in MeOH and treated with an ethereal solution of CH_2N_2 at room temperature for 30 min. After evaporation of the solvent from the reaction mixture, the dimethyl ester (**1a**) was crystallized from MeOH as colorless crystals (95 mg), mp, 169°; ¹H-nmr (100 MHz, CDCl₃) δ 0.66, 0.69, 0.75, 0.83, 0.88 (each 3H, s, $5 \times CH_3$), 0.96, 1.09, 1.22 (each 6H, s, $6 \times CH_3$), 2.88 (2H, unresolved dd, H-18 and H-18'), 3.25 (m, H-3, H-3', and H-24), 3.61 (6H, s, $2 \times COOCH_3$), 3.69 (2H, s, H-29'), 5.28 (2H, t, *J*=3.13 Hz, H-12 and H-12'); ¹³C-nmr see Table 1; ms *m/z* 984.8 (0.05, **M**⁺, calcd for C₆₂H₉₆O₉, 984.7), 983.75 (0.05), 982.7 (0.1), 966.8 (1.0, **M**⁺-H₂O), 948.8 (1.2, **M**⁺-2H₂O), 934 (2.0), 907 (0.5), 891 (0.7), 875 (0.8), 776 (1.4), 761 (2.3), 760.69 (4.3, **4**, calcd for C₄₈H₇₂O₇, 760.5), 742 (2, **4**-H₂O), 728 (1.1), 716 (1), 700 (1.2), 684 (1.5), 606 (1.2), 552 (0.5), 520 (1.2), 493 (1.8), 468 (7.8), 455 (4.4), 437 (4.2), 435 (3.6), 409 (8.2), 407 (9.2), 391 (6.8), 379 (3.2), 313 (4.0), 293 (9.4), 292 (2.9), 291 (1.7, **5**), 279 (11.2), 261 (49.9, **6**), 260 (39), 255 (11.2), 247 (75.7, **5**-CO₂), 233 (37, **7**), 203 (50.60), 201 (100, **6**-HCOOCH₃), 189 (54), 188 (34.4, **5**-CO₂+COOCH₃), 187 (84.2, **5**-CO₂+HCOOCH₃), 173 (57.2), 161 (42.2), 145 (54.7), 135 (47.5), 133 (55.4), 131 (51.6), 119 (61.9), 105 (60.8), 95 (74.3), 81 (77.6), 54.7 (57). *Anal.* calcd for C₆₂H₉₆O₉·3H₂O; C, 71.63; H, 9.89. Found: C, 71.8; H, 9.6.

ACETYLATION OF **1a**.—Compound **1a** (30 mg) was treated with Ac₂O (15 ml) in pyridine (1 ml) at room temperature overnight. Ice was added to the reaction mixture, whereby a white precipitate was obtained. The precipitate was filtered, washed with H₂O, and dried. It was crystallized from MeOH as white crystals of dimethyl ester triacetate **1b** (25 mg), mp, 157°; ¹H-nmr (100 MHz, CDCl₃); δ 0.72 (6H, s, 2×CH₃), 0.86 (6H, s, 2×CH₃), 0.94 (3H, s, 1×CH₃), 0.98 (3H, s, 1×CH₃), 1.01 (3H, s, 1×CH₃), 1.13 (6H, s, 2×CH₃), 1.25 (3H, s, 1×CH₃), 1.27 (3H, s, 1×CH₃), 2.03, 2.04, 2.05 (each 3H, s, 3×OCOCH₃), 2.9 (2H, dd, *J*=13.75, 5 Hz, H-18 and H-18'), 3.63 (6H, s, 2×COOCH₃), 3.73 (2H, s, H-29'), 4.25 (2H, ABq, *J*=12.5 Hz, H-24), 4.5 (2H, t, *J*=8 Hz, H-3, H-3'), 5.3 (2H, t, *J*=3.25 Hz, H-12 and H-12'); no molecular ion, fragmentation at *m*/z 1050 (1.5 M⁺-AcOH+H), 991 (1.5, M⁺-2AcOH+H), 931 (1.2 M⁺-3AcOH+2H), 866 (0.5), 800 (2), 742 (3), 683 (0.5), 603 (2), 511 (2), 495 (2), 481 (2), 451 (10), 391 (5), 375 (3), 313 (5), 292 (5), 279 (6), 273 (6), 261 (22), 247 (42), 201 (96), 187 (100), 173 (74), 159 (68), 145 (68), 133 (54), 119 (54), 104 (47), 95 (40), 81 (44), 67 (38), 57 (78).

ALKALINE HYDROLYSIS OF 1.—Compound 1 (200 mg) was refluxed with 10% (w/v) NaOH in 70% MeOH, for 4 h; the MeOH from this reaction mixture was evaporated and acidified with HCl. The reaction mixture was precipitated by adding H₂O. The precipitate was filtered and washed with H₂O. It showed two spots on tlc (silica gel; solvent a). It was chromatographed on a silica gel column which was eluted with a solvent gradient of increasing polarity of C₆H₆ and EtOAc to afford 2 and 3.

 $_{3\beta,29}$ -DIHYDROXYOLEAN-12-EN-28-OIC ACID (**2**).—The fractions eluted with C₆H₆-EtOAc (1:1) yielded white crystals on evaporation which were then recrystallized from MeOH as colorless sharp needles (120 mg); mp, $_{302}^{\circ}$ [α]D +70.42 (C=0.3, MeOH) lit. mp, $_{305-309}^{\circ}$, [α]D +70.7; ir max (KBr) 3420 (OH), 2930 (CH₂), 1695 (CO, acid), 1465, 1385, 1200, 1045, 1030, 825 (s) (trisubstituted double bond) cm⁻¹; ms m/z 472 (3, M⁺), 455 (3), 441 (2), 316 (2), 264 (48, RDA, **8**) 233 (100, **8**-CH₂OH), 219

(10), 207 (23, RDA, 9), 201 (46), 187 (11), 175 (8), 159 (10), 147 (8), 133 (8), 119 (14), 105 (13), 95 (12), 81 (14), 69 (14), 49 (16). *Anal.* calcd for: $C_{30}H_{48}O_4$: C, 76.32; H, 10.23. Found: C, 76.20; H, 10.40.

METHYLATION OF 2.—Compound 2 (50 mg) was methylated as 1 to yield the monomethyl ester (2a). It was crystallized from MeOH as colorless needles (48 mg), mp, 210°, lit. mp, 213.5-214.5°; ms m/z 486 (3, M⁺), 455 (6), 278 (16, RDA, 8a), 247 (100, 8a-CH₂OH), 207 (16, RDA, 9), 201 (34). The ¹H-nmr data are identical as those published in literature (2).

ACETYLATION OF **2a**.—Compound **2a** (20 mg) was acetylated and worked up as described above in the case of **1** to yield monomethyl ester diacetate (**2b**). It was crystallized from MeOH as fine colorless crystals (15 mg) mp, 225°; ms m/z 570 (3, M⁺), 510 (7), 451 (2), 320 (30, RDA, **8b**), 247 (100, **8b**-CH₂OAc), 201 (74).

3B,24-DIHYDROXYOLEAN-12-EN-28,29-DIOIC ACID (**3**).—Compound **3** was isolated from the fractions eluted with C_6H_6 -EtOAc (1:4) and purified by recrystallization from MeOH as small colorless crystals (50 mg), mp, 290°, [α]D +66.66 (C=0.03, MeOH), uv λ max (MeOH) 210 nm; ir ν max (KBr) 3425 (OH), 2925 (C-H), 1692 (CO, acid), 1470, 1380, 1255, 1120, 1045, 1025, 825 (s) (trisubstituted double bond) cm⁻¹; ms m/z 502 (1, M⁺), 466 (1, M⁺-2H₂O), 453 (1, M⁺-H₂O-CH₂OH), 425 (1), 412 (1, M⁺-2COOH), 394 (1, 412-H₂O), 379 (2), 340 (1), 332 (2), 316 (1), 286 (2), 278 (100, RDA, **10**), 260 (9), 233 (35, **10**-COOH), 223 (15, RDA, **11**), 207 (16), 187 (**10** -2COOH+H), 189 (18, 233-CO₂), 175 (46, 189-CH₂), 159 (18), 144 (13), 131 (11), 119 (19), 105 (28), 92 (18), 81 (22), 69 (12), 55 (22).

METHYLATION OF (3).—Compound 3 (40 mg) was methylated in the usual manner as described above to yield the dimethyl ester (3a), which was crystallized from MeOH as colorless crystals (35 mg), mp, 200°, $[\alpha]D + 30.3$ (C=0.07 CHCl₃); ¹H-nmr (100 MHz, CDCl₃), $\delta 0.66$ (3H, s, 26-Me), 0.83 (3H, s, 25-Me), 1.10 (3H, s, 27-Me), 1.22 (6H, s, 23 and 30 Me), 2.88 (1H, dd, J=5, 13.75 Hz, H-18), 3.35 (m, H-3 and H-24), 3.60, 3.65 (each 3H, s, 2×COOCH₃), 5.29 (1H, t, J=3.75 Hz, H-12); ms m/z 530 (5, M⁺), 512 (1, M⁺ -H₂O), 499 (2, M⁺ -CH₂OH), 481 (2), 471 (5, M⁺ -COOCH₃), 453 (1), 393 (1), 360 (1), 306 (50, RDA, 10a), 301 (1), 292 (10), 274 (4), 259 (2), 247 (46, 10a-COOCH₃), 233 (12, 247-CH₂), 223 (10, RDA, 11), 215 (22), 205 (12), 188 (18, 10a-2 COOCH₃), 187 (100, 10a-2COOCH₃+H), 175 (28), 174 (22), 159 (12), 144 (9), 131 (10), 119 (12), 105 (16), 92 (15), 81 (18), 69 (10), 55 (17).

ACETYLATION OF **3a**.—Compound **3a** (15 mg) was acetylated as **1** to yield **3b**, which was crystallized from MeOH as white crystals of dimethyl ester diacetate (**3b**) (12 mg), mp, 141°; ¹H-nmr (100 MHz, CDCl₃), δ 0.71 (3H, s, 26-Me), 0.93 (3H, s, 23-Me), 1.00 (3H, s, 25-Me), 1.12 (3H, s, 27-Me), 1.24 (3H, s, 30-Me), 2.02, 2.04 (each 3H, s, 2×OCOCH₃), 2.9 (1H, dd, J=5, 13.75 Hz), 3.60, 3.66 (each 3H, s, 2×COOCH₃), 4.25 (2H, ABq, J=12.5 Hz, H-24), 4.57 (1H, t, J=8 Hz, H-3), 5.3 (1H, t, J=3.75 Hz, H-12); ms m/z 614 (1, M⁺), 554 (6, M⁺-AcOH+H), 495 (2, 554-COOCH₃), 435 (4, 495-AcOH+H), 307 (14, RDA, **11a**), 306 (36, RDA, **10a**), 292 (18, **10a**-CH₂), 247 (47, **10a**-COOCH₃), 232 (14, 247-CH₃), 215 (24), 187 (100, **10a**-2COOCH₃+H), 174 (22), 119 (12), 81 (14), 55 (10).

ATTEMPTED ESTERIFICATION OF 2 WITH 3.—A mixture (40 mg) of compound 2 and 3 was refluxed with methanolic HCl (10 ml MeOH, 1 ml H₂O, 1.7 ml HCl) for 5 h. The MeOH from this reaction mixture was evaporated and H₂O added, which furnished a white precipitate. It was filtered and dried. It showed identical spots (silica gel, solvent a) with that of the original mixture of 2 and 3. No spot corresponding to 1 was detected.

ACKNOWLEDGMENTS

The authors thank the University Grants Commission of Pakistan for financial support of this work.

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Received 25 April 1984